

Propensity scores for multiple treatments: A tutorial for the `mnps` function in the `twang` package

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1 Introduction

The Toolkit for Weighting and Analysis of Nonequivalent Groups, `twang`, was designed to make causal estimates in the binary treatment setting. In `twang` versions 1.3 and later, we have extended this software package to handle more than two treatment conditions through the new `mnps` function, which stands for multinomial propensity scores. McCaffrey et al. (2013) describe the methodology behind the `mnps` function; the purpose of this document is to describe the syntax and features related to the implementation in `twang`.

At a high level, the `mnps` function decomposes the propensity score estimation into several applications of the `ps` function, which was designed for the standard dichotomous treatment setting. For this reason, users who are new to `twang` are encouraged to learn about the `ps` function before using the `mnps` function. The other vignette that accompanies the package (Ridgeway et al., 2012) provides an extensive overview of the `ps` function, and much of that information will not be repeated here.

2 An ATE example

To demonstrate the package we utilize a random subset of the data described in McCaffrey et al. (2013). This truncated dataset is called `AOD`, and is included in the package. There are three treatment groups in the study, and the data include records for 200 youths in each treatment group of an alcohol and other drug treatment evaluation. We begin by loading the package and the data. Because there is a stochastic component to the subsequent model fits, we also set the random seed to ensure full replicability.

```
> library(twang)
> data(AOD)
> set.seed(1)
```

For the `AOD` dataset, the variable `treat` contains the treatment indicators, which have possible values `community`, `metcbt5`, and `scy`. The other variables included in the dataset are:

- `suf12`: outcome variable, substance use frequency at 12 month follow-up

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- `illact`: pretreatment covariate, illicit activities scale
- `crimjust`: pretreatment covariate, criminal justice involvement
- `subprob`: pretreatment covariate, substance use problem scale
- `subdep`: pretreatment covariate, substance use dependence scale
- `white`: pretreatment covariate, indicator for non-Hispanic white youth

In such an observational study, there are several quantities that one may be interested in estimating. The estimands that are most commonly of interest are the average treatment effect on the population (ATE) and the average treatment effect on the treated (ATT). The differences between these quantities are explained at length in McCaffrey et al. (2013), but in brief the ATE answers the question of how, on average, the outcome of interest would change if everyone in the population of interest had been assigned to a particular treatment relative to if they had all received another single treatment. The ATT answers the question of how the average outcome would change if everyone who received one particular treatment had instead received another particular treatment.

The main argument for the `mnps` function is a formula with the treatment variable on the left-hand side of a tilde, and pre-treatment variables on the right-hand side, separated by plus signs. Other key arguments are `data`, which simply tells the function the name of the dataframe that contains the variables for the propensity score estimation; the `estimand`, which can either be “ATT” or “ATE”; and `verbose`, which if set as `TRUE` instructs the function to print updates on the model fitting process, which can take a few minutes.

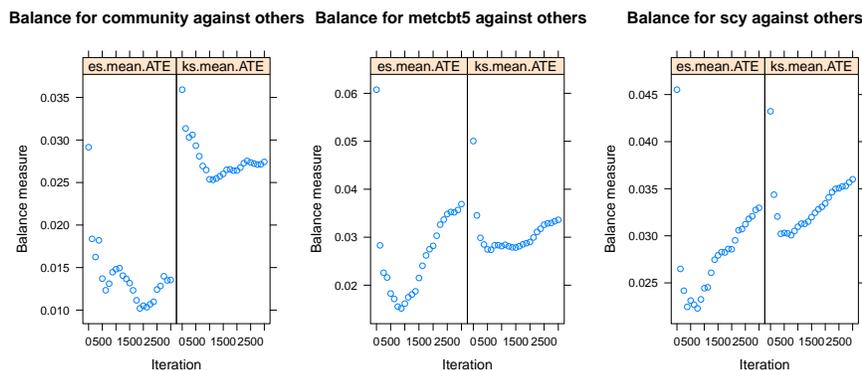
```
> mnps.AOD <- mnps(treat ~ illact + crimjust + subprob + subdep + white,
+                 data = AOD, estimand = "ATE", verbose = FALSE,
+                 stop.method = c("es.mean", "ks.mean"),
+                 n.trees = 3000)
```

The `twang` methods rely on tree-based regression models that are built in an iterative fashion. As the iterations or number of regression trees added to the model increases, the model becomes more complex. However, at some point, more complex models typically result in worse balance on the pre-treatment variables and therefore are less useful in a propensity score weighting context. The `n.trees` argument controls the maximum number of iterations.

Another key choice is the measure of balance that one uses when fitting these models. This is specified in the `stop.method` argument. As with the `ps` function, four `stop.method` objects are included in the package. They are `es.mean`, `es.max`, `ks.mean`, and `ks.max`. The four stopping rules are defined by two components: a balance metric for covariates and rule for summarizing across covariates. The balance metric summarizes the difference between two univariate distributions of a single pre-treatment variable (e.g., illicit activities scale). The default stopping rules in `twang` use two balance metrics: absolute standardized bias (also referred to as the absolute standardized mean difference or the effect size (ES)) and the Kolmogorov-Smirnov (KS) statistic. The stopping rule use two different rules for summarizing across covariates: the mean of the covariate balance metrics (“mean”) or the maximum of the balance metrics (“max”). The first piece of the stopping rule name identifies the balance metric (ES or KS) and the second piece specifies the method for summarizing across balance metrics. For instance, `es.mean` uses the effect size or the absolute standardized bias and summarizes across variables with the mean and the `ks.max` uses the KS statistics to assess balances and summarizes using the maximum across variables and the other two stopping rules use the remaining two combinations of balance metrics and summary statistics. In this example, we chose to examine both `es.mean` and `ks.mean`, which is the default.

A first step is to make sure that we let the models run for a sufficiently large number of iterations in order to optimize the balance statistics of interest. We do this by seeing whether any of the balance measures of interest still appear to be decreasing after the number of iterations specified by the argument `n.trees` (10,000 iterations is the default).

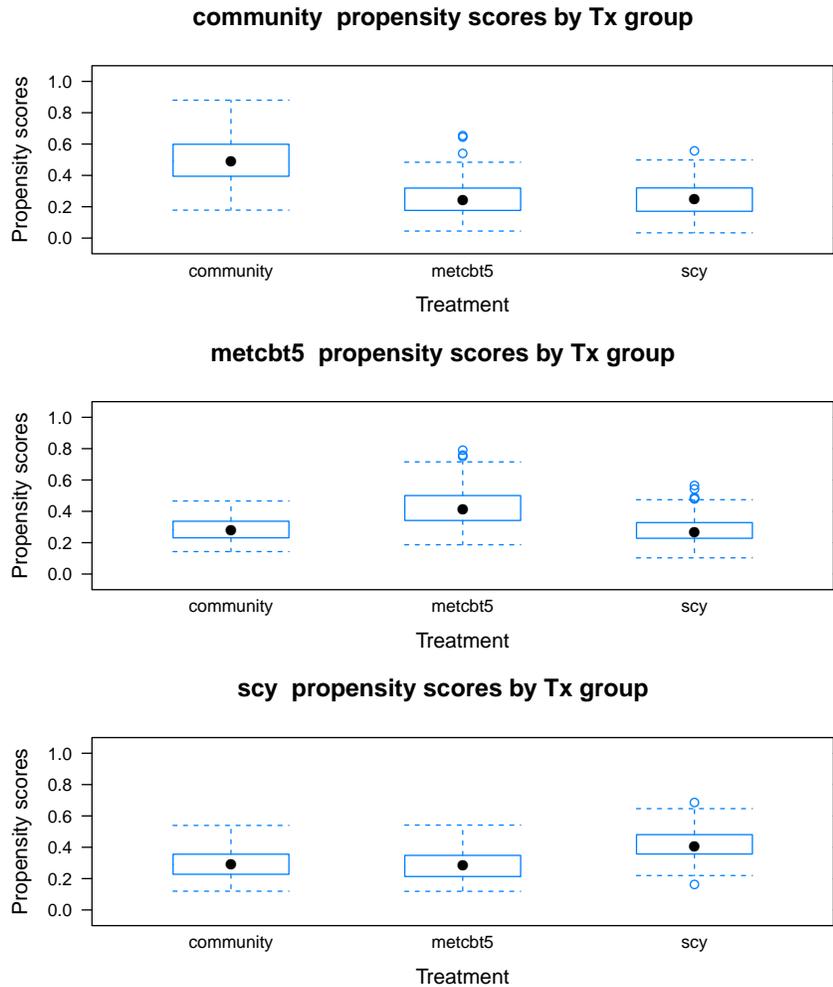
```
> plot(mnps.AOD, plots = 1)
```



In this figure, it appears that each of the balance measures are optimized with substantially fewer than 3,000 iterations, so we do not have evidence that we should re-run the `mnps()` call with a higher number of iterations or trees.

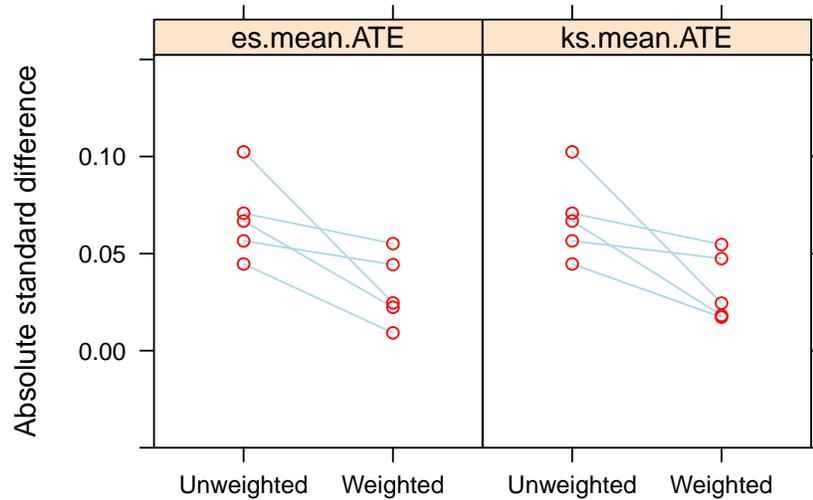
A key assumption in propensity score analyses is that each experimental unit has a non-zero probability of receiving each treatment. The plausibility of this assumption may be assessed by the overlap of the empirical propensity score distributions. This diagnostic is available using the `plots = 2` argument in the `plot` function. Here, the overlap assumption generally seems to be met, although there should be some concern that adolescents in the `metcibt5` and `scy` conditions do not overlap well with the community group given the top most graphic. See McCaffrey et al. (2013) for more details on this issue.

```
> plot(mnps.AOD, plots = 2)
```



As with the `ps` function for the binary treatment setting, the default plotting function for `mnps`-class objects also displays information on commonly-used balance statistics. In particular, it provides comparisons of the absolute standard differences (setting the `plots` argument equal to 3) and t statistics (with the `plots` argument equal to 4), before and after weighting. However, whereas there is a single plot for these balance diagnostics in the binary treatment setting, in the multiple treatment case, one can either examine a plot for each of the treatment conditions, or summarize the balance statistics in some way, across the treatment conditions. As a default, the `plot` function for an `mnps` object returns the maximum of the balance statistics across treatment groups for each of the covariates:

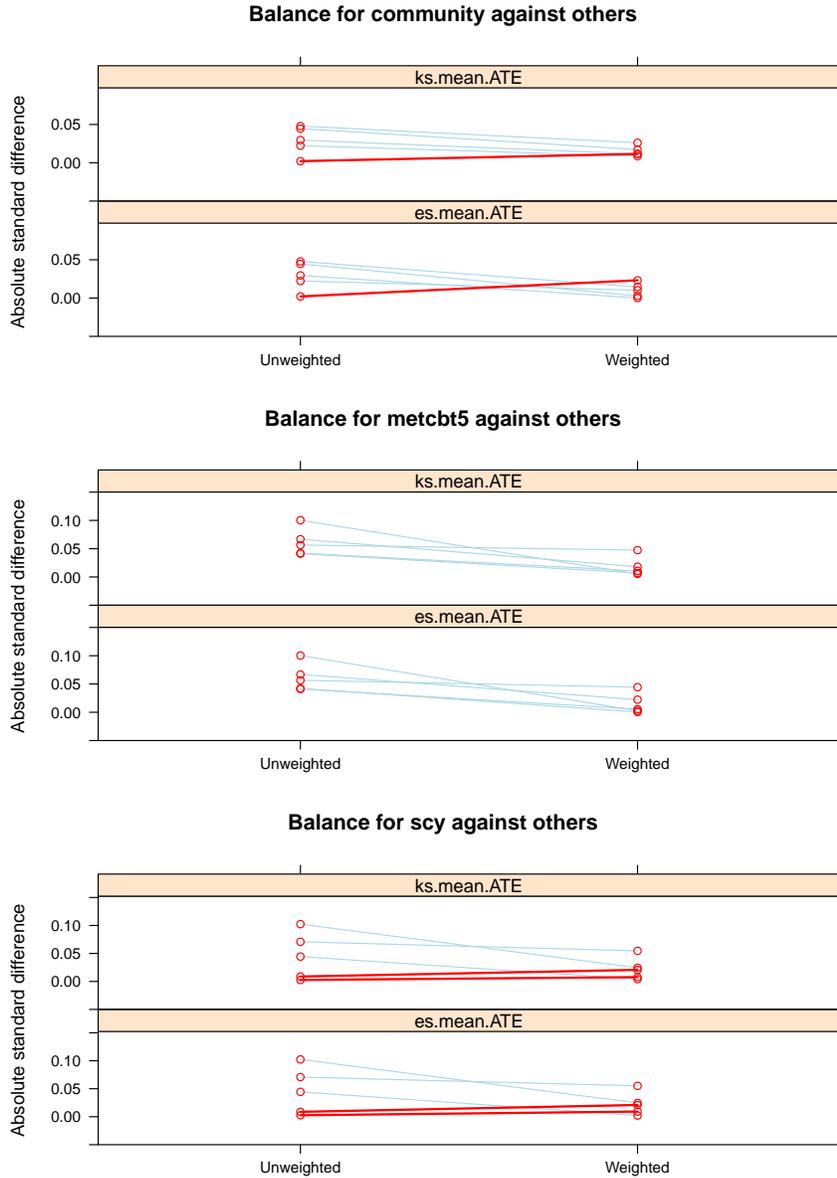
```
> plot(mnps.AOD, plots = 3)
```



If any of the differences had been statistically significant (before taking the maximum across treatment groups), the corresponding hollow circles in this plot would be solid.

It is possible to adjust the summarizing function using the `summaryFcn` argument. For example, one might consider the mean absolute standard differences rather than the maximum by setting `summaryFcn = mean`. Note that the function name should be provided without quotes. Regardless of the summary function, the circles at the end of the line segments will be hollow if none of the differences is statistically significant, and will be solid if at least one is significant. Another useful option is setting that argument equal to `NULL` which avoids the summary step altogether, and displays the balance statistics for each of the treatment conditions separately:

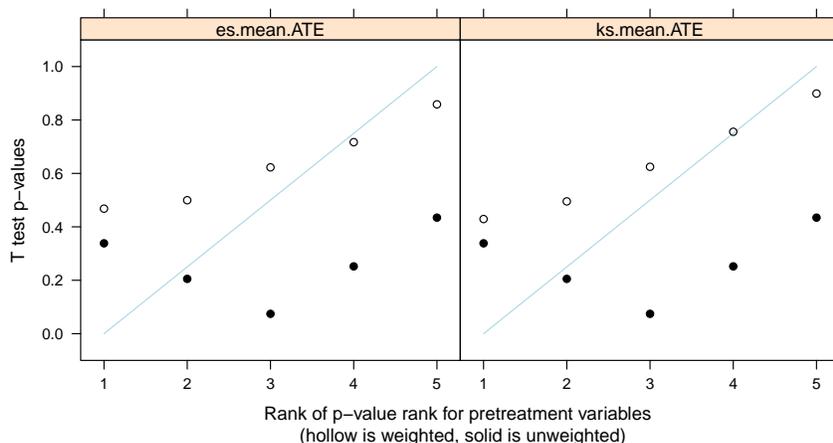
```
> plot(mnps.AOD, plots = 3, summaryFcn = NULL, figureRows = 3)
```



The additional `figureRows` argument instructs the function to spread the plots over three rows; by default the plots would be arranged in a single row rather than a column.

Setting the `plots` argument equal to 4 compares weighted and unweighted t -test or χ^2 statistic p -values for differences between each of the individual treatment groups and observations in all other treatment groups. Note that KS p -values are not available for ATE in the multiple treatment setting, and the `plots` argument therefore may not be set to 5.

```
> plot(mnps.AOD, plots = 4)
```



Beyond graphics, there are several other functions that may be of interest to `mnps` users. The first is `means.table` which provides a nice, simple summary of balance across the groups. When `estimand` is set as 'ATE', the table shows the population means for each pretreatment covariate in the first column as well as each treatment group's unweighted and ATE weighted means and corresponding unweighted and weighted population standardized mean differences. As shown in the table below, incorporation of the ATE propensity score weights improves each treatment groups overall balance with the population means for each pretreatment covariate. The function also includes an argument called `includeSD` whose default is `FALSE`; changing it to `TRUE` returns standard deviations for each of the treatment conditions (not shown).

```
> means.table(mnps.AOD, stop.method = "es.mean", digits = 3)
```

	pop.mean	unwt.community.mean	wt.community.mean	unwt.community.smd	
illact	0.075	0.097	0.085	0.022	
crimjust	-0.068	-0.065	-0.092	0.002	
subprob	-0.016	-0.060	-0.013	-0.045	
subdep	0.015	0.046	0.015	0.030	
white	0.178	0.160	0.173	-0.048	
	wt.community.smd	unwt.metcbt5.mean	wt.metcbt5.mean	unwt.metcbt5.smd	
illact	0.010	0.007	0.052	-0.067	
crimjust	-0.023	0.037	-0.065	0.100	
subprob	0.003	0.026	-0.016	0.042	
subdep	0.000	0.058	0.021	0.041	
white	-0.015	0.200	0.195	0.057	
	wt.metcbt5.smd	unwt.scy.mean	wt.scy.mean	unwt.scy.smd	wt.scy.smd
illact	-0.022	0.120	0.077	0.044	0.002
crimjust	0.003	-0.174	-0.093	-0.102	-0.025
subprob	0.000	-0.013	-0.007	0.003	0.009
subdep	0.005	-0.058	-0.042	-0.071	-0.055
white	0.044	0.175	0.170	-0.009	-0.021

More extensive balance information is given by the `bal.table` function. For propensity score analyses with multiple treatments, this function returns a lot of information. For each outcome

category, and each stopping rule (in addition to the unweighted analysis) the `bal.table` function gives balance statistics such as weighted and unweighted means by treatment group. Note in this case that the “control” columns (labeled `ct`) refer to every treatment group except the one that is considered the treatment for a particular output. For example, in the first table that follows, `tx` refers to `community` treatment, and `ct` refers to all treatments except `community`.

```
> bal.table(mnps.AOD)
```

```
$community
```

```
$community$unw
```

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.097	1.045	0.075	1.014	0.022	0.382	0.703	0.037	NA
crimjust	-0.065	1.050	-0.068	1.041	0.002	0.036	0.971	0.038	NA
subprob	-0.060	0.965	-0.016	0.985	-0.045	-0.782	0.434	0.058	NA
subdep	0.046	1.079	0.015	1.031	0.030	0.501	0.617	0.028	NA
white	0.160	0.368	0.178	0.383	-0.048	-0.847	0.397	0.018	NA

```
$community$es.mean.ATE
```

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.085	1.001	0.075	1.014	0.010	0.139	0.889	0.024	NA
crimjust	-0.092	1.018	-0.068	1.041	-0.023	-0.254	0.799	0.036	NA
subprob	-0.013	0.938	-0.016	0.985	0.003	-0.178	0.858	0.044	NA
subdep	0.015	1.046	0.015	1.031	0.000	0.083	0.934	0.023	NA
white	0.173	0.379	0.178	0.383	-0.015	-0.462	0.644	0.006	NA

```
$community$ks.mean.ATE
```

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.083	1.001	0.075	1.014	0.009	0.136	0.892	0.025	NA
crimjust	-0.080	1.018	-0.068	1.041	-0.011	-0.193	0.847	0.031	NA
subprob	0.001	0.948	-0.016	0.985	0.017	-0.023	0.982	0.032	NA
subdep	0.003	1.052	0.015	1.031	-0.012	-0.031	0.975	0.027	NA
white	0.168	0.375	0.178	0.383	-0.026	-0.577	0.564	0.010	NA

```
$metc5
```

```
$metc5$unw
```

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.007	1.035	0.075	1.014	-0.067	-1.147	0.252	0.065	NA
crimjust	0.037	1.038	-0.068	1.041	0.100	1.743	0.082	0.077	NA
subprob	0.026	1.019	-0.016	0.985	0.042	0.716	0.474	0.047	NA
subdep	0.058	1.047	0.015	1.031	0.041	0.709	0.478	0.042	NA
white	0.200	0.401	0.178	0.383	0.057	0.958	0.338	0.022	NA

```
$metc5$es.mean.ATE
```

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.052	0.997	0.075	1.014	-0.022	-0.363	0.717	0.038	NA
crimjust	-0.065	1.003	-0.068	1.041	0.003	0.283	0.777	0.029	NA
subprob	-0.016	0.997	-0.016	0.985	0.000	0.062	0.950	0.028	NA
subdep	0.021	1.037	0.015	1.031	0.005	0.085	0.932	0.030	NA
white	0.195	0.397	0.178	0.383	0.044	0.726	0.468	0.017	NA

```
$metcbt5$ks.mean.ATE
      tx.mn tx.sd  ct.mn ct.sd std.eff.sz  stat    p    ks ks.pval
illact  0.056 0.999  0.075 1.014   -0.018 -0.311 0.756 0.036    NA
crimjust -0.073 1.008 -0.068 1.041   -0.005  0.178 0.859 0.026    NA
subprob  -0.026 1.000 -0.016 0.985   -0.011 -0.067 0.947 0.025    NA
subdep   0.022 1.034  0.015 1.031    0.007  0.097 0.923 0.032    NA
white    0.197 0.398  0.178 0.383    0.047  0.791 0.429 0.018    NA
```

```
$scy
$scy$unw
      tx.mn tx.sd  ct.mn ct.sd std.eff.sz  stat    p    ks ks.pval
illact  0.120 0.963  0.075 1.014    0.044  0.790 0.430 0.057    NA
crimjust -0.174 1.028 -0.068 1.041   -0.102 -1.788 0.074 0.062    NA
subprob  -0.013 0.972 -0.016 0.985    0.003  0.045 0.964 0.037    NA
subdep  -0.058 0.964  0.015 1.031   -0.071 -1.268 0.205 0.058    NA
white    0.175 0.381  0.178 0.383   -0.009 -0.151 0.880 0.003    NA
```

```
$scy$es.mean.ATE
      tx.mn tx.sd  ct.mn ct.sd std.eff.sz  stat    p    ks ks.pval
illact  0.077 0.992  0.075 1.014    0.002  0.052 0.958 0.033    NA
crimjust -0.093 1.010 -0.068 1.041   -0.025 -0.492 0.623 0.036    NA
subprob  -0.007 0.973 -0.016 0.985    0.009  0.149 0.881 0.029    NA
subdep  -0.042 0.973  0.015 1.031   -0.055 -0.675 0.500 0.044    NA
white    0.170 0.377  0.178 0.383   -0.021 -0.427 0.669 0.008    NA
```

```
$scy$ks.mean.ATE
      tx.mn tx.sd  ct.mn ct.sd std.eff.sz  stat    p    ks ks.pval
illact  0.079 0.992  0.075 1.014    0.004  0.071 0.943 0.033    NA
crimjust -0.093 1.011 -0.068 1.041   -0.024 -0.489 0.625 0.036    NA
subprob  -0.008 0.972 -0.016 0.985    0.008  0.127 0.899 0.029    NA
subdep  -0.041 0.974  0.015 1.031   -0.055 -0.682 0.495 0.044    NA
white    0.170 0.377  0.178 0.383   -0.021 -0.419 0.675 0.008    NA
```

Finally, there is also `summary` method for `mnps` objects which gives some information on balance measures as well as the number of iterations (trees) selected for each model under each stopping rule.

```
> summary(mnps.AOD)
```

```
Summary of mnps object:
```

```
Summary of community against others.
```

```
      n.treat n.ctrl ess.treat ess.ctrl  max.es  mean.es  max.ks
unw          200    400 200.0000 400.0000 0.04785366 0.02930366 0.05833333
es.mean.ATE  200    400 184.5124 389.3660 0.02322317 0.01018732 0.04353362
ks.mean.ATE  200    400 188.3758 392.3995 0.02607402 0.01499874 0.03192988
      max.ks.p  mean.ks iter
unw            NA 0.03600000  NA
es.mean.ATE    NA 0.02639685 1874
ks.mean.ATE    NA 0.02507839 1082
```

Summary of metcbt5 against others.

	n.treat	n.ctrl	ess.treat	ess.ctrl	max.es	mean.es	max.ks
unw	200	400	200.0000	400.0000	0.10027943	0.06137739	0.07666667
es.mean.ATE	200	400	186.1874	394.2979	0.04431685	0.01501965	0.03800035
ks.mean.ATE	200	400	188.3648	395.4704	0.04744407	0.01774854	0.03562304
	max.ks.p	mean.ks	iter				
unw	NA	0.05033333	NA				
es.mean.ATE	NA	0.02830985	880				
ks.mean.ATE	NA	0.02724992	581				

Summary of scy against others.

	n.treat	n.ctrl	ess.treat	ess.ctrl	max.es	mean.es	max.ks
unw	200	400	200.0000	400.0000	0.10238503	0.04578066	0.06166667
es.mean.ATE	200	400	189.5017	393.4114	0.05514367	0.02240795	0.04425948
ks.mean.ATE	200	400	190.0047	393.7790	0.05466257	0.02231721	0.04368363
	max.ks.p	mean.ks	iter				
unw	NA	0.04333333	NA				
es.mean.ATE	NA	0.03013078	754				
ks.mean.ATE	NA	0.02992878	712				

After examining the graphical and tabular diagnostics provided by `twang`, we can analyze the outcome variable using the propensity scores generated by the `mnps` function. Although two stop methods were specified initially (`es.mean` and `ks.mean`), at this point we have to commit to a single set of weights. From the `bal.table` call above, we see that the balance properties are very similar for the two stopping rules, and from the `summary` statement, we see that the effective sample sizes (`ess.treat`) are similar as well. Hence, we expect the two stop methods to give similar results; we choose to analyze the data with the `es.mean` weights.

In order to analyze the data using the weights, it is recommended that one use the `survey` package, which performs weighted analyses. We can add the weights to the dataset using the `get.weights` function and specify the survey design as follows:

```
> require(survey)
> AOD$w <- get.weights(mnps.AOD, stop.method = "es.mean")
> design.mnps <- svydesign(ids=~1, weights=~w, data=AOD)
```

As shown in the `ps` vignette, we can then perform the propensity score-adjusted regression using the `svyglm` function:

```
> glm1 <- svyglm(suf12 ~ as.factor(treat), design = design.mnps)
> summary(glm1)
```

Call:

```
svyglm(formula = suf12 ~ as.factor(treat), design = design.mnps)
```

Survey design:

```
svydesign(ids = ~1, weights = ~w, data = AOD)
```

Coefficients:

	Estimate	Std. Error	t value
(Intercept)	-0.09913	0.06736	-1.472
as.factor(treat)metcbt5	0.14858	0.10502	1.415
as.factor(treat)scy	0.06464	0.09998	0.647

```

                                Pr(>|t|)
(Intercept)                      0.142
as.factor(treat)metcbt5          0.158
as.factor(treat)scy              0.518

```

(Dispersion parameter for gaussian family taken to be 1.002082)

Number of Fisher Scoring iterations: 2

Using this small subset of the data, we are unable to detect differences in the treatment group means. However, the coefficient for the metcbt5 term represents the causal effect of metcbt5 vs. community and the coefficient for the scy term represents the causal effect of scy vs. community assuming there are no unobserved confounders. In the context of this application, the signs of the estimates correspond to higher substance use frequency for youths exposed to metcbt5 or scy relative to community. More details on how to obtain all relevant pairwise differences can be found in McCaffrey et al. (2013).

3 An ATT example

It is also possible to explore treatment effects on the treated (ATTs) using the `mnps` function. A key difference in the multiple treated setting is that we must be clear as to which treatment condition “the treated” refers to. This is done through the `treatATT` argument. Here, we define the treatment group of interest to be the community group; thus, we are trying to draw inferences about the relative effectiveness of the three treatment groups for individuals like those who were enrolled in the community program.

```

> mnps.AOD.ATT <- mnps(treat ~ illact + crimjust + subprob + subdep + white,
+                       data = AOD, estimand = "ATT", treatATT = "community",
+                       verbose = FALSE, n.trees = 3000)

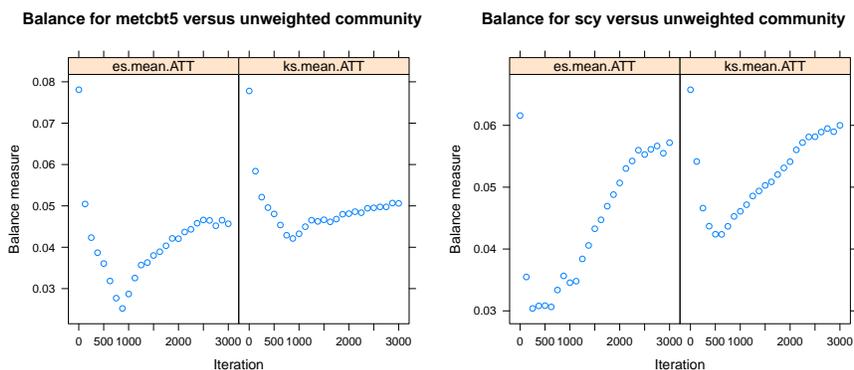
```

The same array of visual and numerical summaries are available as they were in the ATE analysis.

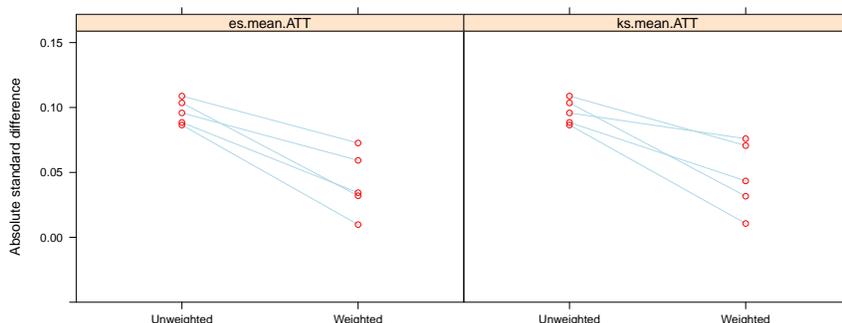
```

> plot(mnps.AOD.ATT, plots = 1)

```



```
> plot(mnps.AOD.ATT, plots = 3)
```



Although the same basic graphical descriptions are available as in the ATE case, note that the population means above are replaced with the means of the `treatATT` category in the `means.table` call.

```
> means.table(mnps.AOD.ATT, digits = 3)
```

	community.mean	unwt.metcbt5.mean	wt.metcbt5.mean	unwt.metcbt5.smd	
illact	0.097	0.007	0.086	0.087	
crimjust	-0.065	0.037	-0.032	-0.097	
subprob	-0.060	0.026	-0.062	-0.088	
subdep	0.046	0.058	0.057	-0.011	
white	0.160	0.200	0.186	-0.109	
	wt.metcbt5.smd	unwt.scy.mean	wt.scy.mean	unwt.scy.smd	wt.scy.smd
illact	0.011	0.120	0.098	-0.021	-0.001
crimjust	-0.032	-0.174	-0.041	0.104	-0.023
subprob	0.002	-0.013	-0.018	-0.048	-0.043
subdep	-0.011	-0.058	-0.036	0.096	0.076
white	-0.071	0.175	0.163	-0.041	-0.008

The `bal.table` output is similar to the ATE case. However, for ATT, we do not need tables that give balance for the `treatATT` category against itself.

```
> bal.table(mnps.AOD.ATT)
```

Note that ``tx`` refers to the category specified as the `treatATT`, `community`.

```
$metcbt5
$metcbt5$unw
```

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.097	1.045	0.007	1.035	0.087	0.870	0.385	0.100	0.270
crimjust	-0.065	1.050	0.037	1.038	-0.097	-0.980	0.328	0.105	0.221
subprob	-0.060	0.965	0.026	1.019	-0.088	-0.861	0.390	0.090	0.394
subdep	0.046	1.079	0.058	1.047	-0.011	-0.113	0.910	0.055	0.924
white	0.160	0.368	0.200	0.401	-0.109	-1.041	0.298	0.040	0.997

```
$metcbt5$ks.mean.ATT
```

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
--	-------	-------	-------	-------	------------	------	---	----	---------

illact	0.097	1.045	0.086	1.023	0.011	0.102	0.919	0.042	0.995
crimjust	-0.065	1.050	-0.032	0.997	-0.032	-0.313	0.754	0.051	0.959
subprob	-0.060	0.965	-0.062	0.988	0.002	0.018	0.986	0.039	0.997
subdep	0.046	1.079	0.057	1.048	-0.011	-0.104	0.917	0.050	0.963
white	0.160	0.368	0.186	0.390	-0.071	-0.662	0.509	0.026	1.000

\$metcibt5\$es.mean.ATT

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.097	1.045	0.087	1.024	0.010	0.094	0.925	0.041	0.995
crimjust	-0.065	1.050	-0.032	0.998	-0.032	-0.317	0.752	0.051	0.957
subprob	-0.060	0.965	-0.062	0.989	0.003	0.025	0.980	0.039	0.998
subdep	0.046	1.079	0.058	1.049	-0.012	-0.112	0.911	0.051	0.959
white	0.160	0.368	0.187	0.391	-0.073	-0.680	0.497	0.027	1.000

\$scy

\$scy\$unw

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.097	1.045	0.120	0.963	-0.021	-0.223	0.823	0.060	0.866
crimjust	-0.065	1.050	-0.174	1.028	0.104	1.048	0.295	0.080	0.545
subprob	-0.060	0.965	-0.013	0.972	-0.048	-0.481	0.631	0.090	0.394
subdep	0.046	1.079	-0.058	0.964	0.096	1.012	0.312	0.085	0.466
white	0.160	0.368	0.175	0.381	-0.041	-0.401	0.688	0.015	1.000

\$scy\$ks.mean.ATT

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.097	1.045	0.098	1.036	-0.001	-0.006	0.995	0.050	0.960
crimjust	-0.065	1.050	-0.041	0.973	-0.023	-0.235	0.814	0.039	0.998
subprob	-0.060	0.965	-0.018	0.979	-0.043	-0.402	0.688	0.045	0.987
subdep	0.046	1.079	-0.036	0.994	0.076	0.744	0.457	0.074	0.664
white	0.160	0.368	0.163	0.370	-0.008	-0.077	0.939	0.003	1.000

\$scy\$es.mean.ATT

	tx.mn	tx.sd	ct.mn	ct.sd	std.eff.sz	stat	p	ks	ks.pval
illact	0.097	1.045	0.100	1.005	-0.002	-0.023	0.982	0.056	0.902
crimjust	-0.065	1.050	-0.064	0.995	-0.002	-0.016	0.988	0.052	0.941
subprob	-0.060	0.965	-0.027	0.967	-0.034	-0.336	0.737	0.055	0.904
subdep	0.046	1.079	-0.018	0.993	0.059	0.596	0.551	0.069	0.707
white	0.160	0.368	0.176	0.382	-0.045	-0.433	0.665	0.016	1.000

The process to analyze the outcome variable is also similar:

```

> require(survey)
> AOD$w.ATT <- get.weights(mnps.AOD.ATT, stop.method = "es.mean")
> design.mnps.ATT <- svydesign(ids=~1, weights=~w.ATT, data=AOD)

> glm1 <- svyglm(suf12 ~ as.factor(treat), design = design.mnps.ATT)
> summary(glm1)

```

Call:

```
svyglm(formula = suf12 ~ as.factor(treat), design = design.mnps.ATT)
```

Survey design:

```
svydesign(ids = ~1, weights = ~w.ATT, data = AOD)
```

Coefficients:

	Estimate	Std. Error	t value
(Intercept)	-0.10505	0.06383	-1.646
as.factor(treat)metcbt5	0.20071	0.10409	1.928
as.factor(treat)scy	0.08076	0.09901	0.816
	Pr(> t)		
(Intercept)	0.1003		
as.factor(treat)metcbt5	0.0543		
as.factor(treat)scy	0.4150		

Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1

(Dispersion parameter for gaussian family taken to be 0.9746663)

Number of Fisher Scoring iterations: 2

Note in this case that the estimated treatment effect of community on those exposed to the community treatment is slightly stronger than in the ATE case (high numbers are bad for the outcome variable). Although not statistically significant, such differences are compatible with the notion that the youths who actually received the community treatment responded more favorably to it than the “average” youth would have (where the average is taken across the whole collection of youths enrolled in the study).

The discussion in McCaffrey et al. (2013) may be useful for determining whether the ATE or ATT is of greater interest in a particular application.

4 Conclusion

Often, more than two treatments are available to study participants. If the study is not randomized, analysts may be interested in using a propensity score approach. Previously, few tools existed to aide the analysis of such data, perhaps tempting analysts to ignore all but two of the treatment conditions. We hope that this extension to the `twang` package will encourage more appropriate analyses of observational data with more than two treatment conditions.

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References

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