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Title Continuous and Dichotomized Index Predictors Based on Distribution Quantiles

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Description Select optimal functional regression or dichotomized quantile predictors for survival/logistic/numeric outcome and perform optimistic bias correction for any optimally dichotomized numeric predictor(s), as in Yi, et. al. (2023) <doi:10.1016/j.labinv.2023.100158>.

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Index

Continuous and Dichotomized Index Predictors Based on Distribution Quantiles

Description

Primary functions in this package are

- optimSplit_dichotom() optimal dichotomizing predictor(s) selection via dichotomizing split sample Still need? Select optimal functional regression or dichotomized quantile predictors for survival/logistic/numeric outcome
- BBC_dichotom() Bootstrap-based optimism correction for dichotomizing selected predictor(s) Still need? perform optimism correction for any optimal dichotomizing predictor(s)
- clusterQp() calculate user-selected sample quantiles in each cluster of observations.
- FRindex() Functional regression index as a predictor in the functional regression model

References

Selection of optimal quantile protein biomarkers based on cell-level immunohistochemistry data. Misung Yi, Tingting Zhan, Amy P. Peck, Jeffrey A. Hooke, Albert J. Kovatich, Craig D. Shriver, Hai Hu, Yunguang Sun, Hallgeir Rui and Inna Chervoneva. Under revision

Quantile index biomarkers based on single-cell expression data. Misung Yi, Tingting Zhan, Amy P. Peck, Jeffrey A. Hooke, Albert J. Kovatich, Craig D. Shriver, Hai Hu, Yunguang Sun, Hallgeir Rui and Inna Chervoneva. Laboratory Investigation, 2023. doi:10.1016/j.labinv.2023.100158

BBC_dichotom

Description

Functions explained in this documentation are,

- BBC_dichotom() to obtain a multivariable regression model with bootstrap-based optimism correction on the dichotomized predictors.
- optimism_dichotom() a helper function to compute the bootstrap-based optimism of the dichotomized predictors.
- coef_dichotom() a helper function to obtain the estimated multivariable regression coefficients
 of the dichotomized predictors.

Usage

```
BBC_dichotom(formula, dichotom, data, ...)
```

```
optimism_dichotom(formula, X, data, R = 100L, ...)
```

```
coef_dichotom(formula, dX, data)
```

Arguments

formula	formula, left-hand-side being the response y and right-hand-side being the pre- dictors <i>in addition to</i> the predictors to be dichotomized. If there is no additional predictor, use $y \sim 1$
dichotom	one-sided formula of the set of predictors to be dichotomized. These predictors can be stored in data as one or more numeric columns and/or one matrix column
data	data.frame, containing the response y and predictors in formula, as well as the predictors to be dichotomized
	additional parameters, currently not in use
Х	(for helper function optimism_dichotom()) numeric matrix of k columns, a set of k numeric predictors
R	positive integer scalar, number of bootstrap replicates R , default 100L
dX	(for helper function $coef_dichotom()$) logical matrix of k columns, a set of k dichotomized predictors

Details

Function BBC_dichotom() obtains a multivariable regression model with bootstrap-based optimism correction on the dichotomized predictors. Specifically,

- Dichotomize the k predictors in the *entire data* (using function m_rpartD()). Fit a regression model to the entire data with the k dichotomized predictors as well as the additional predictors, if any (using helper function coef_dichotom()). The estimated regression model is referred to as the *apparent performance*.
- 2. Obtain the bootstrap-based optimism based on R copies of bootstrap samples, using optimism_dichotom. Calculate the median of bootstrap-based optimism, specific to each of the dichotomized predictors. In future, we may expand the options to include the use of trimmed-mean mean.default(, trim), etc. For now, let's refer to the median optimism as the optimism-correction of the k dichotomized predictors.

Subtract the optimism-correction (in Step 2) from the apparent performance estimates (in Step 1), *only for the k dichotomized predictors*. The apparent performance estimates for the additional predictors, if any, are not modified. The variance-covariance (vcov) estimates of the apparent performance is not modified, for now. None of the other regression model diagnostics, such as residuals, logLikelihood, etc., are modified neither, for now. The coefficient-only, partially-modified regression model is referred to as the *optimism-corrected performance*.

Value

Function BBC_dichotom returns a coxph, glm or lm regression model, with attributes,

- attr(,'optimism') the returned object from optimism_dichotom
- attr(,'apparent_cutoff') a double vector, cutoff thresholds for the k predictors in the apparent model

Details of Helper Function optimism_dichotom()

Function optimism_dichotom computes the bootstrap-based optimism of the dichotomized predictors. First, *R* bootstrap samples are generated, for which the end-user may specify a Random seed, if needed. Then,

- 1. From each of the *R* bootstrap samples, obtain the dichotomizing branches for the *k* predictors to be dichotomized, using function m_rpartD()
- Dichotomize the k predictors in each bootstrap sample using the respective dichotomizing branches from Step 1. The regression coefficients estimate for the k dichotomized predictors (using helper function coef_dichotom()) is referred to as the bootstrap performance estimate.
- 3. Dichotomize the k predictors in the *entire data* using each of the bootstrap dichotomizing branches from Step 1. The regression coefficients estimate for the k dichotomized predictors (using helper function coef_dichotom()) is referred to as the *test performance estimate*.

The difference between the bootstrap and test performance estimates, based on each of the R bootstrap samples, are referred to as the bootstrap-based *optimism* or optimistic bias.

Details of Helper Function coef_dichotom()

Function coef_dichotom obtains the estimated multivariable regression coefficients of the dichotomized predictors. A Cox proportional hazards (coxph) regression for Surv response, a logistic (glm) regression for logical response, or a linear (lm) regression for gaussian response is performed with

- the dichotomous logical predictors, given as the columns of dX, and
- the additional predictors specified in formula

When dX has duplicated columns, the regression model is fitted using the *unique* columns of dX and the additional predictors in formula. The returned coefficient estimates repeat the corresponding estimates of the unique columns of dX.

Returns of Helper Functions

Helper function optimism_dichotom() returns an $R \times k$ double matrix of bootstrap-based optimism, with attributes

attr(,'cutoff') an $R \times k$ double matrix, the R copies of bootstrap cutoff thresholds for the k predictors. See attribute 'cutoff' of function m_rpartD()

Helper function coef_dichotom() returns a double vector of the coefficients of the dichotomized predictors, with attributes

attr(,'model') the coxph, glm or lm regression model

References on Helper Function optimism_dichotom()

Ewout W. Steyerberg (2009) Clinical Prediction Models. doi:10.1007/9780387772448

Frank E. Harrell Jr., Kerry L. Lee, Daniel B. Mark. (1996) Multivariable prognostic models: issues in developing models, evaluating assumptions and adequacy, and measuring and reducing errors. doi:10.1002/(SICI)10970258(19960229)15:4<361::AIDSIM168>3.0.CO;24

Examples

```
library(survival)
data(flchain, package = 'survival') # see more details from ?survival::flchain
head(flchain2 <- within.data.frame(flchain, expr = {
    mgus = as.logical(mgus)
}))
dim(flchain3 <- subset(flchain2, futime > 0)) # required by ?rpart::rpart
dim(flchain_Circulatory <- subset(flchain3, chapter == 'Circulatory'))
m1 = BBC_dichotom(Surv(futime, death) ~ age + sex + mgus,
data = flchain_Circulatory, dichotom = ~ kappa + lambda)
summary(m1)
attr(attr(m1, 'optimism'), 'cutoff')
attr(m1, 'apparent_cutoff')
```

celldata

Description

Ki67 cell data containing 622 patients

Usage

Ki67

Format

PATIENT_ID factor, unique patient identifier

tissueID factor, TMA core identifier

RECURRENCE integer, recurrence indicator, 1 = Recurred, 0 = not Recurred

RECFREESURV_MO integer, recurrence-free survival time in months

Marker double, cell signal intensity of the protein immunofloerscence signal

inner_x integer, x-coordinate in the cell centroid in the TMA core

inner_y integer, y-coordinate in the cell centroid in the TMA core

AGE_AT_DX integer, age at diagnosis

Tstage integer, tumor stage

NodeSt integer, node stage, -1 = unknown, 0 = Node Negative, 1 = Node Positive

HRpos integer, indicator of hormone positive status (ER+ or PR+), 1 = positive, 0 = negative

HistologicalGrade integer, histology grade

Her2_path_qIF integer, Her2 status, 1 = positive, 0 = negative

RACE character, race, White, Black, Asian, Native Hawaiian or Other Pacific Islander, American Indian or Alaska Native, Unknown

RadjCHEMO integer, adjuvant chemo treatment, 0 = unknown, 1 = done, 2 = NOT done

- RadjRAD integer, adjuvant radiation treatment, 0 = unknown, 1 = done, 2 = NOT done
- HORM_4cat integer, hormone treatment, 0 = unknown, 1 = not indicated, 2 = done, 3 = recommended, but not done

MSI double, mean signal intensity (mean over all cells in the TMA core)

clusterQp

Description

Obtain vectors of sample quantiles in each cluster of observations

Usage

```
clusterQp(
   formula,
   data,
   exclude,
   from = 0.01,
   to = 0.99,
   by = 0.01,
   type = 7,
   ...
)
```

Arguments

formula	formula passed to aggregate.formula. To calculate the cluster-specific statistics for response y , the user may use
	y ~ id to retain only the cluster id in the returned value
	y ~ id + x1 + x2 to retain the cluster id and cluster-specific variables x_1 and x_2 in the returned value
	y ~ . to retain all (supposedly cluster-specific) variables from data in the re- turned value
data	data.frame
exclude	(optional) formula or character vector, (supposedly non-cluster-specific) variables to be excluded from aggregation. To remove variables z_1 and z_2 , the user may use either
	 exclude = c('z1', 'z2'); or exclude = . ~ z1 - z2
from, to, by	double scalars, the starting, end, and increment values to specify a sequence of probabilities $p = (p_1, \dots, p_N)'$ for the sample quantiles $q = (q_1, \dots, q_N)'$
type	integer scalar, type of quantile algorithm
	additional parameters, currently not in use

Details

Function clusterQp() calculates N sample quantiles in each aggregated cluster of observations. The aggregation is specified by parameters formula and exclude.

Value

Function clusterQp() returns an aggregated data.frame. A double matrix of N columns is created to store the sample quantiles q of each aggregated cluster. The column names of this quantile matrix are the probabilities p.

Examples

```
Ki67q = clusterQp(Marker ~ ., data = Ki67, exclude = c('tissueID','inner_x','inner_y'))
tmp = clusterQp(Marker ~ ., data = Ki67, exclude = . ~ . - tissueID - inner_x - inner_y)
# stopifnot(identical(Ki67q, tmp))
# stopifnot(!anyDuplicated.default(Ki67q$subjID))
head(Ki67q)
sapply(Ki67q, FUN = class)
```

FRindex

Functional Regression Indices & Weights

Description

Functions explained in this documentation are,

- FRindex() to compute the functional regression indices and weights based on the functional predictors.
- predict.FRindex() to compute the predicted values based on functional regression indices and weights model.
- FR_gam() a helper function to fit a functional regression model using generalized additive models with integrated smoothness estimation (gam).

Usage

```
FRindex(formula, data, sign_prob = 0.5, ...)
```

```
FR_gam(
  formula,
  data,
  xarg = as.double(colnames(X)),
  family,
  knot_pct = 0.4,
  knot.value = ceiling(length(xarg) * knot_pct),
  ...
)
## S3 method for class 'FRindex'
predict(
  object,
  newdata = object@data,
```

```
newX = newdata[[object@formula[[3L]]]],
new_xarg = as.double(colnames(newX)),
...
```

Arguments

)

formula	a two-sided formula.
	Left-hand-side is the name of the response <i>y</i> . Supported types of responses are double, logical and Surv.
	Right-hand-side is the name of the tabulated double matrix X of functional predictor values. Each row of X represents the tabulated values for a subject. All rows/subjects are tabulated on a common grid xarg. Each column of X represents the tabulated values at a point on the common grid for each subject.
data	data.frame, with the response y and the tabulated functional predictor values X specified in formula. If the functional predictor is the quantile function, then data is preferably the returned object of clusterQp().
sign_prob	double scalar between 0 and 1, probability corresponding to the selected nearest- even quantile in xarg, which is used to define the sign of the functional regres- sion weights. Default is .5, i.e., the nearest-even median of xarg
	for function predict.FRindex() and helper function FR_gam(), these are currently not in use. For function FRindex(), see a detailed explanation in section Using in FRindex()
xarg	strictly increasing double vector, the common grid on which the functional predictor values X are tabulated
family	<pre>family object, the distribution and link function to be used in gam. Default fam- ily for Surv response is mgcv::cox.ph(), for logical response is binomial(link = 'logit'), for double response is gaussian(link = 'identity').</pre>
knot_pct	positive double scalar, percentage of the number of columns of X , to be used as knot.value. Default is 40%. If knot.value is provided by the end-user, then knot_pct is ignored.
knot.value	positive integer scalar, number of knots (i.e., parameter k in the spline smooth function s) used in gam. Default is the ceiling of knot_pct of the column dimension of X
object	an FRindex object for the predict method, the returned object from function FRindex()
newdata	data.frame, with at least the tabulated functional predictor values X^{new} based on object@formula
newX	double matrix, functional predictor values X^{new} for a set of new subjects. Each row of X^{new} represents the tabulated values for a new subject. All rows/subjects are tabulated on a common grid new_xarg. Each column of X^{new} represents the tabulated values at a point on the common grid for each new subject.
new_xarg	strictly increasing double vector, the common grid on which the functional pre- dictor values X^{new} are tabulated. The length of new_xarg does not need to be the same as the length of object@xarg, but they must share the same range.

Details

Functional regression indices & weights model:

Function FRindex() defines and calculates the functional regression indices and weights in the following steps.

- 1. Fit a functional regression model to the response y using the functional predictor X, with tabulated tabulated on a same grid xarg for all subjects, using helper function FR_gam()
- 2. Select one point in the tabulating grid xarg. For one-dimensional domain, we select the nearest-even quantile of the tabulating grid xarg, corresponding to the user-specified probability sign_prob. Default sign_prob = .5 indicates the median of xarg.
- 3. Obtain the fitted coefficient function $\hat{\beta}(x)$, tabulated on the grid xarg, using internal helper function gam2beta()
- 4. Calculate the integral of the product of the fitted coefficient function $\hat{\beta}(x)$ (from Step 3) and the functional predictor values X, using the trapzoid rule
- 5. Obtain the sign of the correlation between
 - the subject-specific functional predictor *values*, at the selected quantile of xarg (from Step 2), and
 - the subject-specific integrals from Step 4

Functional regression weights (slot @weight) are the tabulated weight function on the grid xarg. These weights are defined as the product of sign (from Step 5) and $\hat{\beta}(x)$ (from Step 3).

Functional regression indices (slot @index) are defined as the product of sign (from Step 5) and intg (from Step 4). Multiplication by sign is required to ensure that the resulting functional regression indices are positively associated with the functional predictor values at the selected quantile of xarg (from Step 2).

Predict method for functional regression indices & weights:

Function predict.FRindex() computes functional regression indices and weights based on the tabulated functional predictors X^{new} in a new sets of subjects. It's important that the new tabulation grid new_xarg must have the same range as the model tabulation grid object@xarg. Then,

- 1. Obtain the fitted coefficient function $\hat{\beta}(x^{new})$ of the existing generalized additive model object@gam, but tabulated on the new grid new_xarg, using internal helper function gam2beta()
- 2. Calculate the integral of the product of the fitted coefficient function $\hat{\beta}(x^{new})$ (from Step 1) and the new functional predictor values X^{new} , using the trapzoid rule

Predicted functional regression weights are the tabulated weight function on the new grid new_xarg. These weights are defined as the product of object@sign and $\hat{\beta}(x^{new})$ (from Step 1).

Predicted functional regression indices are defined as the product of object@sign and intg (from Step 2). Multiplication by object@sign is required to ensure that the resulting functional regression indices are positively associated with the functional predictor values at the selected quantile of object@xarg.

Value

Functional regression indices & weights model:

Function FRindex() returns an S4 FRindex object. The slots of S4 class FRindex are described in section **Slots**.

FRindex

Predict method for functional regression indices & weights:

Function predict.FRindex() returns a double vector, which is the predicted functional regression indices. The returned object contains an attributes

attr(, 'weight') double vector, the predicted functional regression weights

Slots

formula, data, xarg see explanations in section Arguments

gam gam object, the returned object of helper function FR_gam()

- sign double scalar of either 1 or -1, see Step 5 in section Details on function FRindex()
- index,weight double vectors, functional regression indices and functional regression weights, respectively. See section Details on function FRindex()

Using ... in FRindex()

Function FRindex() passes the parameters xarg, family, knot_pct and knot.value into helper function FR_gam() through three dots

The most important parameter among them is xarg. The default argument of the parameter xarg comes from the column names of the matrix of tabulated functional predictor values X. This is particularly convenient when the functional predictor is the quantile function, and data is the returned object of function clusterQp().

Both FRindex() and helper function FR_gam() accept user-provided xarg. In such case, the provided values will be checked such that

- 1. xarg is a numeric vector without missingness
- 2. length of xarg is the same as the number of columns of matrix X
- 3. xarg must be strictly sorted (see is.unsorted)

Otherwise, an error message will be returned.

Details of Helper Function

Helper function FR_gam() uses gam to estimate the functional coefficient by fitting functional regression model.

Returns of Helper Functions

Helper function FR_gam() returns a gam object, with additional attributes

attr(,'X') double matrix of tabulated functional predictor values X

attr(,'xarg') double vector, see explanation of parameter xarg

References

Cui, E., Crainiceanu, C. M., & Leroux, A. (2021). Additive Functional Cox Model. Journal of Computational and Graphical Statistics. doi:10.1080/10618600.2020.1853550

Gellar, J. E., Colantuoni, E., Needham, D. M., & Crainiceanu, C. M. (2015). Cox regression models with functional covariates for survival data. Statistical Modelling. doi:10.1177/1471082X14565526

Examples

```
library(survival)
pt = unique(Ki67$PATIENT_ID)
length(pt) # 622
# set.seed if necessary
train_pt = sample(pt, size = 500L)
Ki67q = clusterQp(Marker ~ ., data = Ki67, exclude = c('tissueID','inner_x','inner_y'))
train_q = subset(Ki67q, PATIENT_ID %in% train_pt)
test_q = subset(Ki67q, !(PATIENT_ID %in% train_pt))
train_q$Marker = log1p(train_q$Marker)
test_q$Marker = log1p(test_q$Marker)
FRi = FRindex(Surv(RECFREESURV_MO, RECURRENCE) ~ Marker, data = train_q)
FRi@index # functional regression index
FRi@weight # functional regression weights
head(show(FRi)) # append `FRi` to the data
(FRi_test = predict(FRi, newdata = test_q))
FRi_train = predict(FRi)
# stopifnot(identical(FRi@index, c(FRi_train)),
# identical(FRi@weight, attr(FRi_train, 'weight')))
# set.seed if necessary
Ki67bbc_v2 = BBC_dichotom(Surv(RECFREESURV_MO, RECURRENCE) ~ NodeSt + Tstage,
  data = data.frame(train_q, FRi_std = std_IQR(FRi_train)),
  dichotom = \sim FRi_std)
summary(Ki67bbc_v2)
Ki67q = clusterQp(Marker ~ ., data = Ki67, exclude = c('tissueID','inner_x','inner_y'))
Ki67q$Marker = log1p(Ki67q$Marker)
library(survival)
FR_gam(Surv(RECFREESURV_MO, RECURRENCE) ~ Marker, data = Ki67q)
```

nlFRindex

Nonlinear Functional Regression Indices

Description

Functions explained in this documentation are,

nlFRindex() to compute the non-linear functional regression indices based on the functional predictors.

predict.FRindex() to compute the predicted values based on functional regression indices model.

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nlFRindex

Usage

```
nlFRindex(
  formula,
  data,
  xarg = as.double(colnames(X)),
  family,
  fit = TRUE,
  ...
)
```

```
## S3 method for class 'nlFRindex'
predict(object, newdata, ...)
```

Arguments

formula	a two-sided formula.
	Left-hand-side is the name of the response <i>y</i> . Supported types of responses are double, logical and Surv.
	Right-hand-side is the name of the tabulated double matrix X of functional predictor values. Each row of X represents the tabulated values for a subject. All rows/subjects are tabulated on a common grid xarg. Each column of X represents the tabulated values at a point on the common grid for each subject.
data	data.frame, with the response y and the tabulated functional predictor values X specified in formula. If the functional predictor is the quantile function, then data is preferably the returned object of clusterQp().
xarg	numeric vector. The default argument comes from the column names of the matrix of tabulated functional predictor values X . This is particularly convenient when the functional predictor is the quantile function, and data is the returned object of function clusterQp(). The user-provided xarg will be checked such that
	1. xarg is a numeric vector without missingness
	2. length of xarg is the same as the number of columns of matrix X
	3. xarg must be strictly sorted (see is.unsorted)
	Otherwise, an error message will be returned.
family	
fit	logical scalar, see gam
	additional parameters, currently not in use
object	an nlFRindex object for the predict method, the returned object from function nlFRindex()
newdata	data.frame, with at least the tabulated functional predictor values X^{new} based on object@formula

Details

Functional regression indices & weights model:

Function nlFRindex() fits a non-linear functional regression model to the response y using the functional predictor X, with values tabulated on a same grid xarg for all subjects (Cui et al, 2021).

Predict method for non-linear functional regression indices:

Function predict.nlFRindex() computes non-linear functional regression indices based on the tabulated functional predictors X^{new} in a new sets of subjects. It's important that the new tabulation grid must be exactly the same as the model tabulation grid object@xarg.

Value

Functional regression indices & weights model:

Function nlFRindex() returns an S4 nlFRindex object. The slots of S4 class nlFRindex are described in section Slots.

Predict method for non-linear functional regression indices:

Function predict.nlFRindex() returns a double vector, which is the predicted non-linear functional regression indices.

Slots

formula, data, xarg see explanations in section Arguments

gam gam object

p.value numeric scalar, p-value for the test of significance of the functional predictor

index double vector, functional regression indices.

References

Cui, E., Crainiceanu, C. M., & Leroux, A. (2021). Additive Functional Cox Model. Journal of Computational and Graphical Statistics. doi:10.1080/10618600.2020.1853550

Examples

```
pt = unique(Ki67$PATIENT_ID)
length(pt) # 622
# set.seed if necessary
train_pt = sample(pt, size = 500L)
Ki67q = clusterQp(Marker ~ ., data = Ki67, exclude = c('tissueID','inner_x','inner_y'))
train_q = subset(Ki67q, PATIENT_ID %in% train_pt)
test_q = subset(Ki67q, !(PATIENT_ID %in% train_pt))
train_q$Marker = log1p(train_q$Marker)
test_q$Marker = log1p(test_q$Marker)
```

```
# using Cox model
m = nlFRindex(Surv(RECFREESURV_MO, RECURRENCE) ~ Marker, data = train_q)
m@p.value # test significance of `Marker` as a functional predictor
train_index = predict(m, newdata = train_q) # non-linear FR index of training data
# stopifnot(identical(train_index, m@index))
```

optimSplit_dichotom

```
predict(m, newdata = test_q) # non-linear FR index of test data
# using logistic regression model
nlFRindex(RECURRENCE ~ Marker, data = train_q)
# using Gaussian model
nlFRindex(RECFREESURV_MO ~ Marker, data = train_q)
```

optimSplit_dichotom Optimal Dichotomizing Predictors via Repeated Sample Splits

Description

Functions explained in this documentation are,

- optimSplit_dichotom() to identify the optimal dichotomizing predictors using repeated sample splits.
- split_dichotom() a helper function to perform a univariable regression model on the test set with a dichotomized predictor, using a dichotomizing rule determined by a recursive partitioning of the training set.

Usage

```
optimSplit_dichotom(formula, data, include, top = 1L, nsplit, ...)
```

split_dichotom(y, x, index, ...)

```
quantile_split_dichotom(y, x, indices = rSplit(y, ...), probs = 0.5, ...)
```

Arguments

formula	formula. Left-hand-side is the name of a Surv, logical, or double response y . Right-hand-side is the candidate numeric predictors in data, given either as the name of a numeric matrix column (e.g., $y \sim X$), or as the names of several numeric vector columns (e.g., $y \sim x1 + x2 + x3$)
data	data.frame, containing the response and predictors in formula
include	language object, inclusion criteria for the optimal dichotomizing predictors. A suggested choice is (highX>.15 & highX<.85) to guarantee a user-desired range of proportions in highX. See explanation of highX in helper function split_dichotom().
top	positive integer scalar, number of optimal dichotomizing predictors, default 1L
nsplit,	additional parameters for function rSplit()

У	(for helper functions) a Surv object, a logical vector, or a double vector, the response y
х	(for helper functions) numeric vector, a single predictor x
index	(for helper function split_dichotom()) logical vector, indices of training and test set. TRUE elements indicate training subjects and FALSE elements indicate test subjects.
indices	(optional, for helper function quantile_split_dichotom()) a list of logical vectors, the indices of multiple training-test sample splits. Default value is provided by function rSplit().
probs	(for helper function quantile_split_dichotom()) double scalar, see quantile

Details

Function optimSplit_dichotom() selects the optimal dichotomizing predictors via repeated sample splits. Specifically,

- 1. Generate multiple training-test sample splits using function rSplit()
- 2. For each candidate predictor, find the median split_dichotom (using helper function quantile_split_dichotom()) of the multiple sample splits from Step 1.
- 3. (Optional) limit the selection in a subset of the candidate predictors. Typically, we would prefer to guarantee a user-desired range of highX (see explanations on highX in section **Returns of Helper Functions**). A suggested choice is (highX>.15 & highX<.85).
- 4. Rank the candidate predictors, from either Step 2 or Step 3, by the decreasing order of the absolute values of the estimated univariable regression coefficients of the corresponding split_dichotom objects.

The *optimal dichotomizing predictors* are the ones with the largest absolute values of the estimated univariable regression coefficients of the corresponding split_dichotom objects.

Value

Function optimSplit_dichotom() returns a data.frame, which contains the response, and only the optimal dichotomizing predictors out of all candidate predictors. Other variables in data, which are not specified in formula, are retained. In addition, the dichotomized values of the optimal dichotomizing predictors, according to their respective dichotomizing rules, are also included. The returned value has attributes,

- attr(,'id_top') positive integer scalar or vector, the indices of the optimal dichotomizing predictors out of all candidate predictors.
- attr(,'top') a diagnostic data.frame of the median split_dichotoms of each of the optimal dichotomizing predictors, with columns
 - \$cutoff the cutoff threshold, identified in the training set
 - \$highX proportion of the dichotomizing predictors greater-than or greater-than-or-equal-to the cutoff threshold, in the test set
 - \$coef the estimated univariable regression coefficient of the dichotomized predictor, in the
 test set

Details on Helper Functions

Univariable regression model with a dichotomized predictor:

Helper function split_dichotom() performs a univariable regression model on the test set with a dichotomized predictor, using a dichotomizing rule determined by a recursive partitioning of the training set. Currently the Cox proportional hazards (coxph) regression for Surv response, logistic (glm) regression for logical response and linear (lm) regression for gaussian response are supported. Specifically, given a training-test sample split,

- 1. find the dichotomizing rule of the response y given the predictor x, using function rpartD(), in the training set
- 2. dichotomize the predictor x using the rule identified in Step 1, in the test set.
- 3. run a univariable regression model on the response y on the dichotomized predictor from Step 2, in the test set.

Quantile of split_dichotom objects:

Helper function quantile_split_dichotom() finds the quantile of the univariable regression coefficient (i.e., effect size) of a dichotomized predictor, based on multiple given training-test sample splits. Specifically,

- 1. for each training-test sample split, fit the univariable regression model based on the dichotomized predictor, using helper function split_dichotom()
- 2. finds the nearest-even (type = 3) quantile of the estimated univariable regression coefficients obtained in Step 1, based on the user-specified probability prob

The split_dichotom object from Step 1, whose estimated univariable regression coefficient equals to the specified quantile identified in Step 2, is referred to as the quantile of split_dichotom objects based on the multiple given training-test sample splits.

Returns of Helper Functions

Helper function split_dichotom(), as well as helper function quantile_split_dichotom(), returns a Cox proportional hazards (coxph), or a logistic (glm), or a linear (lm) regression model, with additional attributes

- attr(,'rule') function, the dichotomizing rule based on the training set
- attr(,'cutoff') numeric scalar, the cutoff threshold based on the training set
- attr(, 'highX') double scalar, proportion of numeric predictor x, in the test set, which is greaterthan or greater-than-or-equal-to the cutoff threshold attr(, 'cutoff')
- attr(,'coef') double scalar, the estimated univariable regression coefficient of the dichotomized predictor in the test set

Examples

```
library(survival)
data(pbc, package = 'survival') # see more details from ?survival::pbc
head(pbc2 <- within.data.frame(subset(pbc, status != 1L), expr = {
    death = (status == 2L)
    trt = structure(trt, levels = c('D-penicillmain', 'placebo'), class = 'factor')
    trt = relevel(trt, ref = 'placebo')</pre>
```

rpartD

```
}))
# set.seed if needed
m1 = optimSplit_dichotom(
   Surv(time, death) ~ bili + chol + albumin + copper + alk.phos + ast + trig + platelet + protime,
   data = pbc2, nsplit = 20L, include = (highX > .15 & highX < .85), top = 2L)
head(m1, n = 10L)
attr(m1, 'top')</pre>
```

rpartD

Dichotomize via Recursive Partitioning

Description

Dichotomize one or more predictors of a Surv, a logical, or a double response, using recursive partitioning and regression tree rpart.

Usage

```
rpartD(
  y,
  x,
  check_degeneracy = TRUE,
  cp = .Machine$double.eps,
  maxdepth = 2L,
  ...
)
m_rpartD(y, X, check_degeneracy = TRUE, ...)
```

Arguments

У	a Surv object, a logical vector, or a double vector, the response y
х	numeric vector, one predictor x
check_degenerac	У
	logical scalar, whether to allow the dichotomized value to be all-FALSE or all-TRUE (i.e., degenerate) for any one of the predictors. Default TRUE to produce a warning message for degeneracy.
ср	double scalar, complexity parameter, see rpart.control. Default .Machine\$double.eps, so that a split is enforced no matter how small improvement in overall R^2 is
maxdepth	positive integer scalar, maximum depth of any node, see rpart.control. Default 2L, because only the first node is needed
	additional parameters of rpart and/or rpart.control
Х	numeric matrix, a set of predictors. Each column of X is one predictor.

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rpartD

Details

Dichotomize Single Predictor:

Function rpartD() dichotomizes one predictor in the following steps,

- 1. Recursive partitioning and regression tree rpart analysis is performed for the response y and the predictor x.
- The labels.rpart of the first node of the rpart tree is considered as the dichotomizing rule of the double predictor x. The term *dichotomizing rule* indicates the combination of an inequality sign (>, >=, < and <=) and a double cutoff threshold a
- 3. The dichotomizing rule from Step 2 is further processed, such that
 - < a is regarded as $\ge a$
 - $\leq a$ is regarded as > a
 - > a and $\ge a$ are regarded as is.

This step is necessary for a narrative of *greater than* or *greater than or equal to* the threshold *a*.

4. A warning message is produced, if the dichotomizing rule, applied to a new double predictor newx, creates an all-TRUE or all-FALSE result. We do not make the algorithm stop, as most regression models in R are capable of handling an all-TRUE or all-FALSE predictor, by returning a NA_real_ regression coefficient estimate.

Dichotomize Multiple Predictors:

Function m_rpartD() dichotomizes each predictor X[, i] based on the response y using function rpartD(). Applying the multiple dichotomizing rules to a new set of predictors newX,

- A warning message is produced, if at least one of the dichotomized predictors is all-TRUE or all-FALSE.
- We do not check if more than one of the dichotomized predictors are identical to each other. We take care of this situation in helper function coef_dichotom()

Value

Dichotomize Single Predictor:

Function rpartD() returns a function, with a double vector parameter newx. The returned value of rpartD(y,x)(newx) is a logical vector with attributes

attr(,'cutoff') double scalar, the cutoff value for newx

Dichotomize Multiple Predictors:

Function $m_rpartD()$ returns a function, with a double matrix parameter newX. The argument for newX must have the same number of columns and the same column names as the input matrix X. The returned value of $m_rpartD(y, X)$ (newX) is a logical matrix with attributes

attr(, 'cutoff') named double vector, the cutoff values for each predictor in newX

Note

In future integer and factor predictors will be supported.

Examples

```
## Dichotomize Single Predictor
data(cu.summary, package = 'rpart') # see more details from ?rpart::cu.summary
with(cu.summary, rpartD(y = Price, x = Mileage, check_degeneracy = FALSE))
(foo = with(cu.summary, rpartD(y = Price, x = Mileage)))
foo(rnorm(10, mean = 24.5))
## Dichotomize Multiple Predictors
library(survival)
data(stagec, package = 'rpart') # see more details from ?rpart::stagec
nrow(stagec) # 146
(foo = with(stagec[1:100,], m_rpartD(y = Surv(pgtime, pgstat), X = cbind(age, g2, gleason))))
foo(as.matrix(stagec[-(1:100), c('age', 'g2', 'gleason')]))
```

rSp1	it
------	----

Random Split Sampling with Stratification

Description

Random split sampling, stratified based on the type of the response.

Usage

```
rSplit(y, nsplit, stratified = TRUE, trainFrac = 0.8, ...)
```

Arguments

У	a double vector, a logical vector, a factor, or a Surv object, response y
nsplit	positive integer scalar, replicates of random splits to be performed
stratified	logical scalar, whether stratification based on response \boldsymbol{y} needs to be implemented, default TRUE
trainFrac	double scalar between 0 and 1, fraction of the training set, default .8
	additional parameters, currently not in use

Details

Function rSplit() performs random split sampling, with or without stratification. Specifically,

- If stratified = FALSE, or if we have a double response y, then split the sample into a training and a test set by ratio trainFrac, without stratification.
- Otherwise, split a Surv response y, stratified by its censoring status. Specifically, split subjects with observed event into a training and a test set with training set fraction trainFrac, and split the censored subjects into a training and a test set with training set fraction trainFrac. Then combine the training sets from subjects with observed events and censored subjects, and combine the test sets from subjects with observed events and censored subjects.

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rSplit

- Otherwise, split a logical response y, stratified by itself. Specifically, split the subjects with TRUE response into a training and a test set with training set fraction trainFrac, and split the subjects with FALSE response into a training and a test set with training set fraction trainFrac. Then combine the training sets, and the test sets, in a similar fashion as described above.
- Otherwise, split a factor response y, stratified by its levels. Specifically, split the subjects in each level of y into a training and a test set by ratio trainFrac. Then combine the training sets, and the test sets, from all levels of y.

Value

Function rSplit() returns a length-nsplit list of logical vectors. In each logical vector, the TRUE elements indicate training subjects and the FALSE elements indicate test subjects.

Note

caTools::sample.split() is not what we need.

See Also

split

Examples

rSplit(y = rep(c(TRUE, FALSE), times = c(20, 30)), nsplit = 3L)

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